

Review Article

Neural circuits of pain and itch processing involved in anterior cingulate cortex

Zhi-Gang He^{1*}, Ding-Yu Zhang^{2*}, San-Guang Liu³, Li Feng¹, Mao-Hui Feng⁴, Hong-Bing Xiang¹

¹Department of Anesthesiology and Pain Medicine, Tongji Hospital, Tongji Medical College, Huazhong University of Science & Technology, Wuhan 430030, P. R. China; ²Intensive Care Unit, Wuhan Medical Treatment Center, No. 1 Yintan Road, Dongxihu District, Wuhan 430023, P. R. China; ³Department of Hepatobiliary Surgery, The Second Hospital, Hebei Medical University, Shijiazhuang 050000, P. R. China; ⁴Department of Oncology, Wuhan Peritoneal Cancer Clinical Medical Research Center, Zhongnan Hospital of Wuhan University, Hubei Key Laboratory of Tumor Biological Behaviors & Hubei Cancer Clinical Study Center, No. 169 Donghu Road, Wuhan 430071, Hubei, P. R. China. *Equal contributors.

Received April 2, 2016; Accepted September 2, 2016; Epub December 15, 2016; Published December 30, 2016

Abstract: Anterior cingulate cortex (ACC) is widely considered to be the most important part of the brain's limbic system. Previous researches have shown that ACC plays a significant role in consciousness and behavior, including emotion, sensory, motor, memory, nociceptive processing, etc. Among these connections, nociceptive processing, especially pain and itch, has always been one of the prior research programs for the field of medicine. In recent years, the development of new techniques and advance in neurobiology has led to our new understanding on the link between ACC and nociceptive processing. In addition, neurophysiologic research has revealed that multiple peptides and neurotransmitters, including opioid peptides, glutamate, serotonin and dopamine, are involved in the mechanisms of modulation of nociception by ACC. In this review, we examine a wealth of recent neuroanatomical and neurobiological research that involves ACC and nociceptive processing, and provide a comprehensive review of the neural circuits of pain and itch processing in ACC, and the glutaminergic and opioidergic circuits are especially discussed.

Keywords: Anterior cingulate cortex, pain, itch, neural circuits, opioidergic circuits

Introduction

Anterior cingulate cortex (ACC) is widely considered to be the complex heterogeneous structure of the brain's limbic system. Anatomically, ACC is a distinct subregion of ventromedial frontal cortex consisting of the cingulate sulcus and gyrus that lie dorsal to the corpus callosum and ventral to the superior frontal gyrus. Based upon cytoarchitecture and functional connectivities, ACC is further dichotomized into a more rostral/ventral affective/emotional division comprising pregenual (pgACC) and subgenual ACC (sgACC), which equivalent to Brodmann area (BA) 24a, 24b, 24c, 25, 32 and 33. And a more dorsal cognitive division comprises anterior and posterior portions of the dorsal ACC (adACC and pdACC), equivalent to BA 24a', 24b', 24c', 24d, 32' and 33 (**Figure 1**) [1-3]. Etkin et al. pointed out that between dorsal-

caudal and ventral-rostral subdivisions of the ACC had regulatory roles in emotional processing, and the former was involved in negative emotion whereas the latter was with respect to generating emotional responses [2]. Hsieh et al. indicated that BA24 of caudal ACC and BA9/32 (medial prefrontal cortex/rostral ACC, mPFC/rACC) were preferentially involved in the affective/evaluative processing of pain [4]. Early ablation studies from patients and animals have shown that ACC plays a significant role in consciousness and behavior [5, 6]. Subsequent functional studies using a variety of approaches, including electrical stimulation, microelectrode recording, positron emission tomography (PET) and functional magnetic resonance imaging (fMRI), provided us a further understanding of the link between ACC and emotion, sensory, motor, memory, nociceptive

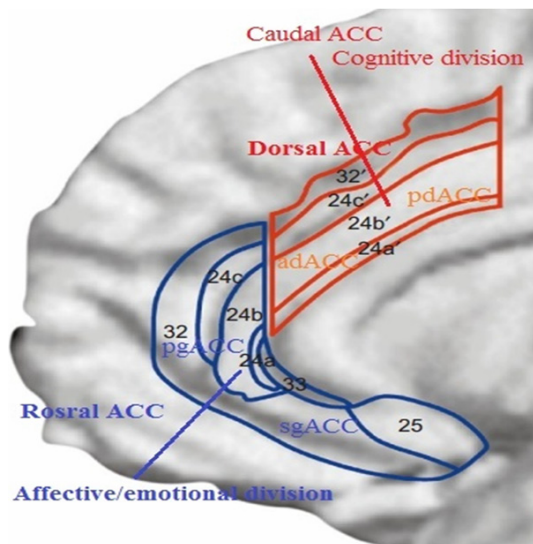


Figure 1. Parcellation of ACC subregions. Abbreviations: sgACC, subgenual ACC; pgACC, pregenual ACC; adACC, anterior dorsal ACC; pdACC, posterior dorsal ACC.

processing [3, 7-10]. Among these connections, nociceptive processing, especially pain and itch, has always been the focus of the research area. Previous reports demonstrated that although pain and itch have some differences, the neural processing of pain has some similarities to that of itch [11-14]. In addition, neurophysiological research has revealed that multiple peptides and neurotransmitters, including opioid peptides, glutamate, serotonin and dopamine, are involved in the modulation of nociception by ACC [15-17]. In this review, we provide a comprehensive review of the neural circuits of pain and itch processing in ACC, and the glutaminergic and opioidergic circuits are especially discussed.

Neural circuit of pain processing in ACC

It has long been known that pain processing is mediated by a neural network consisting of both cortical and subcortical structures. ACC is a key component of the cortical circuit which also includes the primary and secondary somatosensory cortex (SI and SII), the insula and prefrontal regions, whereas the subcortical components of the pain matrix includes the thalamus, amygdala and hippocampus, etc [18, 19]. However, our knowledge about the neural mechanisms of pain processing inside ACC is limited, nor is the neuroanatomical and neurobiological connections between ACC and these

cortical and subcortical cerebral areas. It is generally assumed that the ACC involves the descending pain modulatory system [20]. Study from Freund W et al. indicated that painful electrical stimulation activates ventral posterior ACC more, but during thermal stimulation, there was more activation in dorsal posterior ACC [21]. A research about links between nociception and projections to cortical and motor systems from Bai-Chuang Shyu et al. found that nociceptive responses are generated directly by pyramidal projection neurons in all layers [22]. Davis et al. reported in patients with chronic pain by a positron emission tomography study that there were two distinct ACC areas activated after thalamic stimulation, one in the anterior (aACC) and the other in the posterior portion of the ACC (pACC) [23]. These results suggest that pain processing probably owns a common signaling channel as well as specific neural circuit corresponding to different kinds of pain inside ACC.

The functional connectivity between ACC and other cerebral structures in pain processing is the focus of neuroscience field. It is now believed that the periaqueductal gray (PAG, known as one of the central pain descending modulation systems [24]) receives direct projections from ACC [25]. The neuroanatomical connection between the ACC and PAG has been revealed by iontophoretic injections of horseradish peroxidase in dorsal, lateral and medial areas of PAG [26]. By a fMRI study about the response to an onset (aversion) and offset (reward) of a noxious heat stimulus of humans and rats, Becerra et al. demonstrated the relevance between rACC and the nucleus accumbens (NAc) in affective and motivational aspects of pain [27], suggesting that rACC-NAc circuit is involved in modulating affective and motivational aspects of pain. A series of research indicates that ACC and the insula subserve the affective-motivational component of pain processing [28-30]. Orbitofrontal cortex, insula, NAc, amygdala and PAG are connected together with ACC through the opioid system [31, 32]. Furthermore, the pathophysiology of chronic visceral pain involved the link between ACC and PAG, insula, amygdala, anterior cingulate cortex, orbitofrontal cortex, medial and dorsolateral prefrontal cortex, and parietal cortex [33]. The thalamus is also one of the most frequently activated regions in human pain research [34]. The retrograde tracing study

Neural processing of pain and itch

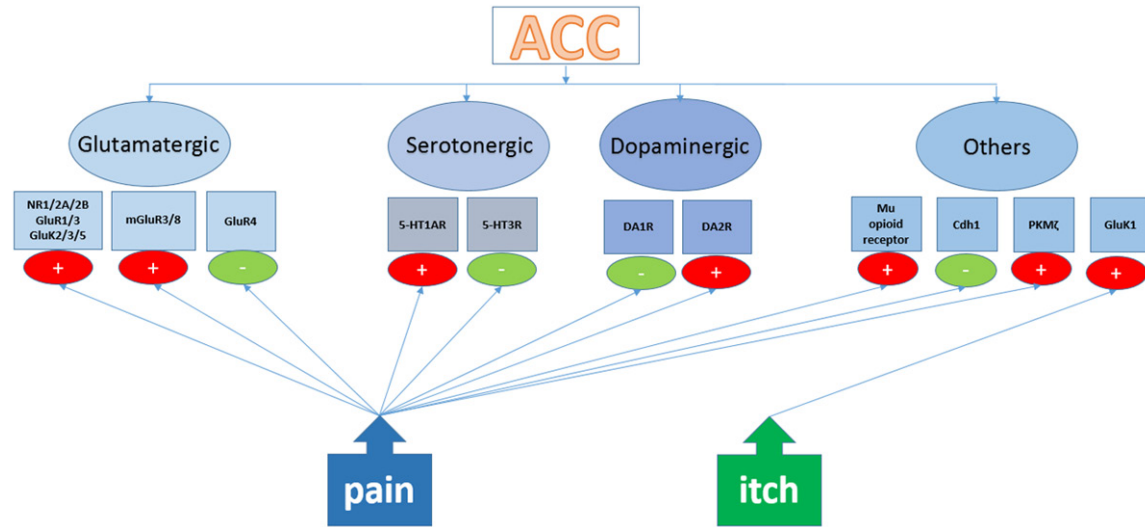


Figure 2. Schematic representation of the signal pathways activated by pain and itch. The expression of mGluR3, mGluR8, GluR1, GluR3, NR1, NR2A, NR2B, 5-HT1AR and mu-opioid receptor in ACC was up-regulated (red oval) whereas GluR4, 5-HT3R, DA1R, DA2R and Cdh1 were down-regulated (blue oval) under pain condition. The expression of PKM ζ in ACC was also up-regulated (red oval). The expression of GluK1 in ACC was up-regulated under itch condition (red oval). “+” and “-” stand for up-regulation and down-regulation, respectively.

from Abrahamson EE et al. demonstrated that the descending projections to the PHA arise from the cingulate cortex and the medial and lateral hypothalamus [35]. Functional tracing of medial nociceptive pathways using MRI observed that noxious stimulation induced enhancement of manganese ion transportation from medial thalamus to cingulate cortex [36]. Neurobiological study indicated that the likely thalamic source of nociceptive inputs to ACC is from the midline, mediodorsal and intralaminar thalamic nuclei (MITN) [37]. These findings suggest that the significance of thalamic-anterior cingulate pathway in pain conditions. In addition, transneuronal tracing with neurotropic pseudorabies viruses (PRV) demonstrated a pathway from ACC to the dorsal hippocampus [38].

The dorsal ACC (dACC) has been shown to be a heterogeneous brain region, which includes two parts, one is a more rostral part of dACC which plays a role in affective processing, and the other is a more caudal region which involves in cognitive and motor processing with projections to the red nucleus and spinal cord [8, 39, 40].

Neurotransmitter and receptor mechanisms underlying ACC

The involvement of the ACC in nociception modulation may be associated with the activities of

a variety of neurotransmitters and peptides, including opioid, glutamate, dopamine and serotonin (Figure 2). Among the three ionotropic glutamate receptor subtypes, researchers have focused their attention most strongly on NMDA receptors. The research about the relationship between endogenous D-serine in the rACC and pain-related negative affect suggested that activation of NMDA receptors in the rACC is necessary for the acquisition of specific pain-related negative emotion [41]. It was reported that painful stimuli potentiate the pre-frontal synaptic transmission and induce glutamate NMDA NR2B receptor expression in the ACC and inhibition of NR2B receptors by administering selective NR2B receptor antagonists locally into the ACC or systemically inhibits inflammation-related allodynia [42-44]. Immunoblot analysis from Yang JX et al. also revealed that chronic constriction injury (CCI) increased the expression of NR2B protein in the ACC and microinjection of NR2B inhibitor ifenprodil into contralateral ACC significantly inhibited CCI-induced thermal hyperalgesia and mechanical allodynia [45]. On the contrary, data from mice with bone cancer pain revealed a significant decrease in the levels of NR1, NR2A, and NR2B subunits of NMDA receptors in the rACC and the results indicated that tumor-induced injury may cause a persistent decrease in NMDA receptor expression in rACC

neurons [46]. Xu H et al. found that both the presynaptic release probability of glutamate and postsynaptic glutamate AMPA receptor-mediated responses were enhanced after injury using the mouse peripheral nerve injury mode, but western blot analysis indicated there was no difference in the expression levels of both GluR1 and GluR2/3 receptors in ACC between the control mice and mice with nerve ligation. Under paclitaxel-induced neuropathic pain (PINP) condition, GluR1, GluR3, GLuK2, GLuK3, GLuK5, NR1 and mGluR8 were significantly up-regulated, whereas GluR2, GLuK1, GLuK4, NR2A and NR2B were not significantly altered and GluR4 was lowly expressed [47]. Recently, Gao SH et al. reported that intra-ACC microinjection of mGluR1-shRNA suppressed the CCI-induced behavioral hypersensitivity, particularly thermal hyperalgesia [48]. Further studies are needed to clarify the effects of various subtypes glutamate receptors in pain processing. The ACC glutamate receptor-based modulation of pain is also associated with other neurotransmitters. The study of López-Avila A et al. suggests an interaction between dopaminergic and glutamatergic systems within the ACC in the genesis and maintenance of long-term nociception [16]. Ortega-Legaspi JM et al. provided evidence of an increase of inhibitory dopaminergic receptor (D2R) mRNA and protein in cg1 in correlation with nociceptive behavior in a neuropathic model of pain in the rat [49]. Ikeda H et al. reported that antagonize of dopamine D1 receptor in the ACC prevented to the analgesic effect of lemon oil [50]. Serotonin (5-HT) and 5-HT receptor are highly involved in pathogenesis of pain [51-62]. Koeppe C et al. found that the activation was significantly reduced after treatment of the 5-HT3 receptor antagonist tropisetron in ACC [63]. Martikainen IK et al. performed psychophysical testing in eleven healthy males who had participated in a positron emission tomography study with [carbonyl-(11C)WAY-100635 ligand for the assessment of 5-HT(1A) receptor binding potential (BP) and demonstrated that 5-HT(1A) receptors in the brain influence pain [17]. Tan W et al. employed spared nerve injury (SNI) model in rat and found down-regulation of Cdh1 protein level as well as up-regulation of AMPA receptor GluR1 subunit protein level in the ACC after SNI surgery and intra-ACC treatment with Cdh1-expressing lentivirus vectors elevated Cdh1 levels and alleviated established

mechanical allodynia in SNI rats [64]. In addition, some research data indicated that PKM ζ in the ACC acts to maintain neuropathic pain [65-67].

The opioidergic circuit in ACC

The ACC is a key region in the processing of pain and the opioid system is bound up with pain. In the early 1990s, it was reported that ACC has one of the highest densities of opioid receptors in the CNS and it has been implicated in acute and chronic pain responses [68]. And there is already growing evidence, on the basis of recent research [69, 70], that the ACC opioid system probably take center stage in processing negative pain affect. However, little is known about opioid system organization and actions.

Different division corresponds to different opioid architectures inside ACC. A rodent study about opioid architecture in the perigenual and midcingulate divisions of ACC from Vogt LJ et al. indicated that perigenual and midcingulate cortices have different opioid architectures due to a higher proportion of mu-opioid receptors expressed by afferent axons in areas 24 and 32 [15]. Further study identified the pgACC as one brain region with a major impact on opioidergic pain modulation [71]. Bruehl S et al. reported that inadequate endogenous opioid inhibitory activity in the rostral anterior cingulate (rACC) contributes to the links between trait anger-out and pain [72]. Interestingly, study on placebo analgesia suggested that the endogenous opioid system is closely linked with it and provides evidence that the rACC represents a crucial cortical area for this type of endogenous pain control [73-76]. A recent research about asymmetry of the endogenous opioid system in ACC suggested that region-specific lateralization of neuronal networks expressing opioid peptides underlies in part lateralization of higher functions, including positive and negative emotions and pain in the human brain [77].

Neural mechanisms of itch processing in ACC

Itch is the most prevalent symptom of allergic and inflammatory skin disease [78, 79]. Although it is known that itch induces activation of a neural network in the brain, the neuro-anatomical pathway of the network as well as

the pathophysiology and neurobiology are not well understood. As a key component of pain processing, ACC also involves itch processing. Imaging study of central itch modulation using PET found that the significant increases of regional cerebral blood flow caused by histamine stimuli were observed in ACC [80, 81]. A series of fMRI study about histamine-mediated and non-histaminergic forms of itch observed signal intensity changes of ACC [82-87]. As with pain experience, the affective and emotional aspects of itch processing are also significantly represented in the activation of ACC, amygdala, and NAc [88-90]. In the field of molecular biology, Descalzi G et al. combined pharmacological, genetic, and electrophysiological approaches and showed that cortical GluK1-containing kainate (KA) receptors are involved in scratching induced by histamine and non-histamine-dependent itching stimuli [91]. Further neuroanatomical and neurobiological studies elucidating the nature of the neural circuits are needed.

Summary

Neuroimaging, neuropathological, and lesion analysis data show that the anterior cingulate cortex is an important cortex region whose central role is to regulate the neural circuits of pain and itch processes. Further in-depth studies are required to explore the an extended anatomical network formed by the neural projections of the ACC in the regulation of the emotional expression and experience of pain or itch.

Acknowledgements

This project was supported by a grant from a Project of China National Natural Science Foundation of PR China (81072152, 81670240), Natural Science Foundation of Hubei Province (2015CFA027), Research Foundation of Health and Family Planning Commission of Hubei Province (WJ2015M), Hebei Province Higher Educational Science and Technology Program, and Health and Family Planning Commission of Hubei Province (No. WJ2015MBO08). The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Mao-Hui Feng, Department of Oncology, Wuhan Peritoneal Cancer Clinical Medical Research Center, Zhongnan Hospital of Wuhan University, Hubei Key Laboratory of Tumor Biological Behaviors & Hubei Cancer Clinical Study Center, Wuhan, Hubei, P. R. China. E-mail: fengmh5690@163.com

References

- [1] Vogt BA and Palomero-Gallagher N. Cingulate gyrus. In: Juergen K Mai, Paxinos G, editors. *The Human Nervous System*. 3rd edition 2012. pp. 943-987.
- [2] Etkin A, Egner T and Kalisch R. Emotional processing in anterior cingulate and medial prefrontal cortex. *Trends Cogn Sci* 2011; 15: 85-93.
- [3] Bush G, Luu P and Posner MI. Cognitive and emotional influences in anterior cingulate cortex. *Trends Cogn Sci* 2000; 4: 215-222.
- [4] Hsieh JC, Meyerson BA and Ingvar M. PET study on central processing of pain in trigeminal neuropathy. *Eur J Pain* 1999; 3: 51-65.
- [5] Lewin W and Whitty CW. Effects of anterior cingulate stimulation in conscious human subjects. *J Neurophysiol* 1960; 23: 445-447.
- [6] Kennard MA. Effect of bilateral ablation of cingulate area on behaviour of cats. *J Neurophysiol* 1955; 18: 159-169.
- [7] Pardo JV, Pardo PJ, Janer KW and Raichle ME. The anterior cingulate cortex mediates processing selection in the Stroop attentional conflict paradigm. *Proc Natl Acad Sci U S A* 1990; 87: 256-259.
- [8] Devinsky O, Morrell MJ and Vogt BA. Contributions of anterior cingulate cortex to behaviour. *Brain* 1995; 118: 279-306.
- [9] Vogt BA, Finch DM and Olson CR. Functional heterogeneity in cingulate cortex: the anterior executive and posterior evaluative regions. *Cereb Cortex* 1992; 2: 435-443.
- [10] Bush G, Frazier JA, Rauch SL, Seidman LJ, Whalen PJ, Jenike MA, Rosen BR and Biederman J. Anterior cingulate cortex dysfunction in attention-deficit/hyperactivity disorder revealed by fMRI and the Counting Stroop. *Biol Psychiatry* 1999; 45: 1542-1552.
- [11] Handwerker HO, Forster C and Kirchhoff C. Discharge patterns of human C-fibers induced by itching and burning stimuli. *J Neurophysiol* 1991; 66: 307-315.
- [12] Jinks SL and Carstens E. Superficial dorsal horn neurons identified by intracutaneous histamine: chemonociceptive responses and modulation by morphine. *J Neurophysiol* 2000; 84: 616-627.
- [13] Schmelz M, Schmidt R, Bickel A, Handwerker HO and Torebjork HE. Specific C-receptors for

Neural processing of pain and itch

- itch in human skin. *J Neurosci* 1997; 17: 8003-8008.
- [14] Drzezga A, Darsow U, Treede RD, Siebner H, Frisch M, Munz F, Weilke F, Ring J, Schwaiger M and Bartenstein P. Central activation by histamine-induced itch: analogies to pain processing: a correlational analysis of O-15 H2O positron emission tomography studies. *Pain* 2001; 92: 295-305.
- [15] Vogt LJ, Sim-Selley LJ, Childers SR, Wiley RG and Vogt BA. Colocalization of mu-opioid receptors and activated G-proteins in rat cingulate cortex. *J Pharmacol Exp Ther* 2001; 299: 840-848.
- [16] Lopez-Avila A, Coffeen U, Ortega-Legaspi JM, del Angel R and Pellicer F. Dopamine and NMDA systems modulate long-term nociception in the rat anterior cingulate cortex. *Pain* 2004; 111: 136-143.
- [17] Martikainen IK, Hirvonen J, Kajander J, Hagelberg N, Mansikka H, Nagren K, Hietala J and Pertovaara A. Correlation of human cold pressor pain responses with 5-HT(1A) receptor binding in the brain. *Brain Res* 2007; 1172: 21-31.
- [18] Price DD. Psychological and neural mechanisms of the affective dimension of pain. *Science* 2000; 288: 1769-1772.
- [19] Treede RD, Kenshalo DR, Gracely RH and Jones AK. The cortical representation of pain. *Pain* 1999; 79: 105-111.
- [20] Kucyi A, Salomons TV and Davis KD. Mind wandering away from pain dynamically engages antinociceptive and default mode brain networks. *Proc Natl Acad Sci U S A* 2013; 110: 18692-18697.
- [21] Freund W, Wunderlich AP, Stuber G, Landwehrmeyer B and Klug R. Graded cutaneous electrical vs thermal stimulation in humans shows different insular and cingulate cortex activation. *Somatosens Mot Res* 2010; 27: 15-27.
- [22] Shyu BC, Sikes RW, Vogt LJ and Vogt BA. Nociceptive processing by anterior cingulate pyramidal neurons. *J Neurophysiol* 2010; 103: 3287-3301.
- [23] Davis KD, Taub E, Duffner F, Lozano AM, Tasker RR, Houle S and Dostrovsky JO. Activation of the anterior cingulate cortex by thalamic stimulation in patients with chronic pain: a positron emission tomography study. *J Neurosurg* 2000; 92: 64-69.
- [24] Mayer DJ. Analgesia produced by electrical stimulation of the brain. *Prog Neuropsychopharmacol Biol Psychiatry* 1984; 8: 557-564.
- [25] Yu R, Gollub RL, Spaeth R, Napadow V, Wasan A and Kong J. Disrupted functional connectivity of the periaqueductal gray in chronic low back pain. *Neuroimage Clin* 2014; 6: 100-108.
- [26] Marchand JE and Hagino N. Afferents to the periaqueductal gray in the rat. A horseradish peroxidase study. *Neuroscience* 1983; 9: 95-106.
- [27] Becerra L, Navratilova E, Porreca F and Borsook D. Analogous responses in the nucleus accumbens and cingulate cortex to pain onset (aversion) and offset (relief) in rats and humans. *J Neurophysiol* 2013; 110: 1221-1226.
- [28] Rainville P, Duncan GH, Price DD, Carrier B and Bushnell MC. Pain affect encoded in human anterior cingulate but not somatosensory cortex. *Science* 1997; 277: 968-971.
- [29] Singer T, Seymour B, O'Doherty J, Kaube H, Dolan RJ and Frith CD. Empathy for pain involves the affective but not sensory components of pain. *Science* 2004; 303: 1157-1162.
- [30] Gu X and Han S. Attention and reality constraints on the neural processes of empathy for pain. *Neuroimage* 2007; 36: 256-267.
- [31] Eippert F, Bingel U, Schoell E, Yacubian J and Buchel C. Blockade of endogenous opioid neurotransmission enhances acquisition of conditioned fear in humans. *J Neurosci* 2008; 28: 5465-5472.
- [32] Pecina M, Azhar H, Love TM, Lu T, Fredrickson BL, Stohler CS and Zubieta JK. Personality trait predictors of placebo analgesia and neurobiological correlates. *Neuropsychopharmacology* 2013; 38: 639-646.
- [33] Fukudo S. Stress and visceral pain: focusing on irritable bowel syndrome. *Pain* 2013; 154 Suppl 1: S63-70.
- [34] Mobascher A, Brinkmeyer J, Warbrick T, Musso F, Wittsack HJ, Saleh A, Schnitzler A and Winterer G. Laser-evoked potential P2 single-trial amplitudes covary with the fMRI BOLD response in the medial pain system and interconnected subcortical structures. *Neuroimage* 2009; 45: 917-926.
- [35] Abrahamson EE and Moore RY. The posterior hypothalamic area: chemoarchitecture and afferent connections. *Brain Res* 2001; 889: 1-22.
- [36] Yang PF, Chen DY, Hu JW, Chen JH and Yen CT. Functional tracing of medial nociceptive pathways using activity-dependent manganese-enhanced MRI. *Pain* 2011; 152: 194-203.
- [37] Hatanaka N, Tokuno H, Hamada I, Inase M, Ito Y, Imanishi M, Hasegawa N, Akazawa T, Nambu A and Takada M. Thalamocortical and intracortical connections of monkey cingulate motor areas. *J Comp Neurol* 2003; 462: 121-138.
- [38] Prasad JA and Chudasama Y. Viral tracing identifies parallel disynaptic pathways to the hippocampus. *J Neurosci* 2013; 33: 8494-8503.
- [39] Russo JF and Sheth SA. Deep brain stimulation of the dorsal anterior cingulate cortex for

- the treatment of chronic neuropathic pain. *Neurosurg Focus* 2015; 38: E11.
- [40] Boccard SG, Fitzgerald JJ, Pereira EA, Moir L, Van Hartevelt TJ, Kringelbach ML, Green AL and Aziz TZ. Targeting the affective component of chronic pain: a case series of deep brain stimulation of the anterior cingulate cortex. *Neurosurgery* 2014; 74: 628-635; discussion 635-627.
- [41] Ren WH, Guo JD, Cao H, Wang H, Wang PF, Sha H, Ji RR, Zhao ZQ and Zhang YQ. Is endogenous D-serine in the rostral anterior cingulate cortex necessary for pain-related negative affect? *J Neurochem* 2006; 96: 1636-1647.
- [42] Zhao MG, Ko SW, Wu LJ, Toyoda H, Xu H, Quan J, Li J, Jia Y, Ren M, Xu ZC and Zhuo M. Enhanced presynaptic neurotransmitter release in the anterior cingulate cortex of mice with chronic pain. *J Neurosci* 2006; 26: 8923-8930.
- [43] Wu LJ, Toyoda H, Zhao MG, Lee YS, Tang J, Ko SW, Jia YH, Shum FW, Zerbinatti CV, Bu G, Wei F, Xu TL, Muglia LJ, Chen ZF, Auberson YP, Kaang BK and Zhuo M. Upregulation of fore-brain NMDA NR2B receptors contributes to behavioral sensitization after inflammation. *J Neurosci* 2005; 25: 11107-11116.
- [44] Chen L, Liu JC, Zhang XN, Guo YY, Xu ZH, Cao W, Sun XL, Sun WJ and Zhao MG. Down-regulation of NR2B receptors partially contributes to analgesic effects of Gentiopicroside in persistent inflammatory pain. *Neuropharmacology* 2008; 54: 1175-1181.
- [45] Yang JX, Hua L, Li YQ, Jiang YY, Han D, Liu H, Tang QQ, Yang XN, Yin C, Hao LY, Yu L, Wu P, Shao CJ, Ding HL, Zhang YM and Cao JL. Caveolin-1 in the anterior cingulate cortex modulates chronic neuropathic pain via regulation of NMDA receptor 2B subunit. *J Neurosci* 2015; 35: 36-52.
- [46] Chiou CS, Huang CC, Liang YC, Tsai YC and Hsu KS. Impairment of long-term depression in the anterior cingulate cortex of mice with bone cancer pain. *Pain* 2012; 153: 2097-2108.
- [47] Masocha W. Astrocyte activation in the anterior cingulate cortex and altered glutamatergic gene expression during paclitaxel-induced neuropathic pain in mice. *Peer J* 2015; 3: e1350.
- [48] Gao SH, Wen HZ, Shen LL, Zhao YD and Ruan HZ. Activation of mGluR1 contributes to neuronal hyperexcitability in the rat anterior cingulate cortex via inhibition of HCN channels. *Neuropharmacology* 2016; 105: 361-377.
- [49] Ortega-Legaspi JM, de Gortari P, Garduno-Gutierrez R, Amaya MI, Leon-Olea M, Coffeen U and Pellicer F. Expression of the dopaminergic D1 and D2 receptors in the anterior cingulate cortex in a model of neuropathic pain. *Mol Pain* 2011; 7: 97.
- [50] Ikeda H, Takasu S and Murase K. Contribution of anterior cingulate cortex and descending pain inhibitory system to analgesic effect of lemon odor in mice. *Mol Pain* 2014; 10: 14.
- [51] Xiang HB, Liu C, Liu TT and Xiong J. Central circuits regulating the sympathetic outflow to lumbar muscles in spinally transected mice by retrograde transsynaptic transport. *Int J Clin Exp Pathol* 2014; 7: 2987-2997.
- [52] Xu AJ, He ZG, Xia XH and Xiang HB. Anesthetic management for craniotomy in a patient with massive cerebellar infarction and severe aortic stenosis: a case report. *Int J Clin Exp Med* 2015; 8: 11534-11538.
- [53] Ye DW, Liu C, Tian XB and Xiang HB. Identification of neuroanatomic circuits from spinal cord to stomach in mouse: retrograde trans-neuronal viral tracing study. *Int J Clin Exp Pathol* 2014; 7: 5343-5347.
- [54] Feng L, Liu TT, Ye DW, Qiu Q, Xiang HB and Cheung CW. Stimulation of the dorsal portion of subthalamic nucleus may be a viable therapeutic approach in pharmacoresistant epilepsy: A virally mediated transsynaptic tracing study in transgenic mouse model. *Epilepsy Behav* 2014; 31: 114-116.
- [55] Ke B, Liu TT, Liu C, Xiang HB and Xiong J. Dorsal subthalamic nucleus electrical stimulation for drug/treatment-refractory epilepsy may modulate melanocortineric signaling in astrocytes. *Epilepsy Behav* 2014; 36: 6-8.
- [56] Liu TT, Feng J, Bu HL, Liu C, Guan XH and Xiang HB. Stimulation for the compact parts of pedunculo-pontine nucleus: An available therapeutic approach in intractable epilepsy. *Epilepsy Behav* 2013; 29: 252-253.
- [57] Liu C, Ye DW, Guan XH, Li RC, Xiang HB and Zhu WZ. Stimulation of the pedunculo-pontine tegmental nucleus may affect renal function by melanocortineric signaling. *Med Hypotheses* 2013; 81: 114-116.
- [58] Liu TT, Guo QQ, An K, Zhang Y, Tian XB, Li RC, Xiang HB and Wang P. The optimal acupoint for acupuncture stimulation as a complementary therapy in pediatric epilepsy. *Epilepsy Behav* 2014; 31: 387-389.
- [59] Pan XC, Song YT, Liu C, Xiang HB and Lu CJ. Melanocortin-4 receptor expression in the rostral ventromedial medulla involved in modulation of nociception in transgenic mice. *J Huazhong Univ Sci Technol Med Sci* 2013; 33: 195-198.
- [60] Qiu Q, Li RC, Ding DF, Liu C, Liu TT, Tian XB, Xiang HB and Cheung CW. Possible mechanism of regulating glucose metabolism with subthalamic nucleus stimulation in parkinson's disease: a virally mediated trans-synaptic tracing study in transgenic mice.

- Parkinsonism Relat Disord 2014; 20: 468-470.
- [61] Liu BW, He ZG, Shen SE and Xiang HB. CeA-RVMM serotonergic circuits and sudden unexpected death in epilepsy. *Int J Clin Exp Med* 2016; 9: 9752-9758.
- [62] Liu BW, Liu QQ, Liu SG and Xiang HB. Renal disease and neural circuits: brain-kidney cross-talk. *Int J Clin Exp Med* 2016; 9: 5326-5333.
- [63] Koeppe C, Schneider C, Thieme K, Mense S, Stratz T, Muller W and Flor H. The influence of the 5-HT₃ receptor antagonist tropisetron on pain in fibromyalgia: a functional magnetic resonance imaging pilot study. *Scand J Rheumatol Suppl* 2004; 119: 24-27.
- [64] Tan W, Yao WL, Hu R, Lv YY, Wan L, Zhang CH and Zhu C. Alleviating neuropathic pain mechanical allodynia by increasing Cdh1 in the anterior cingulate cortex. *Mol Pain* 2015; 11: 56.
- [65] Li XY, Ko HG, Chen T, Descalzi G, Koga K, Wang H, Kim SS, Shang Y, Kwak C, Park SW, Shim J, Lee K, Collingridge GL, Kaang BK and Zhuo M. Alleviating neuropathic pain hypersensitivity by inhibiting PKMzeta in the anterior cingulate cortex. *Science* 2010; 330: 1400-1404.
- [66] Chen Y, Xin Y, Liu C, Chen Y and Cao Y. [Changes of protein kinases Mzeta expression in the anterior cingulate cortex after applying three different magnitude of orthodontic force]. *Zhonghua Kou Qiang Yi Xue Za Zhi* 2014; 49: 748-752.
- [67] Xin Y, Liu X, Cao Y, Chen Y and Liu C. Up-regulation of PKMzeta expression in the anterior cingulate cortex following experimental tooth movement in rats. *Arch Oral Biol* 2014; 59: 749-755.
- [68] Vogt BA, Wiley RG and Jensen EL. Localization of Mu and delta opioid receptors to anterior cingulate afferents and projection neurons and input/output model of Mu regulation. *Exp Neurol* 1995; 135: 83-92.
- [69] Zubieta JK, Smith YR, Bueller JA, Xu Y, Kilbourn MR, Jewett DM, Meyer CR, Koeppe RA and Stohler CS. Regional mu opioid receptor regulation of sensory and affective dimensions of pain. *Science* 2001; 293: 311-315.
- [70] LaGraize SC, Borzan J, Peng YB and Fuchs PN. Selective regulation of pain affect following activation of the opioid anterior cingulate cortex system. *Exp Neurol* 2006; 197: 22-30.
- [71] Sprenger T, Henriksen G, Valet M, Platzer S, Berthele A and Tolle TR. [Positron emission tomography in pain research. From the structure to the activity of the opiate receptor system]. *Schmerz* 2007; 21: 503-513.
- [72] Bruehl S, Burns JW, Chung OY and Chont M. Pain-related effects of trait anger expression: neural substrates and the role of endogenous opioid mechanisms. *Neurosci Biobehav Rev* 2009; 33: 475-491.
- [73] Petrovic P, Kalso E, Petersson KM and Ingvar M. Placebo and opioid analgesia—imaging a shared neuronal network. *Science* 2002; 295: 1737-1740.
- [74] Bingel U, Lorenz J, Schoell E, Weiller C and Buchel C. Mechanisms of placebo analgesia: rACC recruitment of a subcortical antinociceptive network. *Pain* 2006; 120: 8-15.
- [75] Qiu YH, Wu XY, Xu H and Sackett D. Neuroimaging study of placebo analgesia in humans. *Neurosci Bull* 2009; 25: 277-282.
- [76] Murray D and Stoessl AJ. Mechanisms and therapeutic implications of the placebo effect in neurological and psychiatric conditions. *Pharmacol Ther* 2013; 140: 306-318.
- [77] Watanabe H, Fitting S, Hussain MZ, Kononenko O, Iatsyshyna A, Yoshitake T, Kehr J, Alkass K, Druid H, Wadensten H, Andren PE, Nylander I, Wedell DH, Krishtal O, Hauser KF, Nyberg F, Karpyak VM, Yakovleva T and Bakalkin G. Asymmetry of the endogenous opioid system in the human anterior cingulate: a putative molecular basis for lateralization of emotions and pain. *Cereb Cortex* 2015; 25: 97-108.
- [78] Song Y, Pan X, Liu C and Xiang H. Role of nociceptive arcuate nucleus neurons in chloroquine-induced pruritic behaviors in mice. *J Huazhong Univ Sci Technolog Med Sci* 2012; 32: 919-922.
- [79] Liu C, Liu TT, He ZG, Shu B and Xiang HB. Inhibition of itch-related responses by selectively ablated serotonergic signals at the rostral ventromedial medulla in mice. *Int J Clin Exp Pathol* 2014; 7: 8917-8921.
- [80] Mochizuki H, Tashiro M, Kano M, Sakurada Y, Itoh M and Yanai K. Imaging of central itch modulation in the human brain using positron emission tomography. *Pain* 2003; 105: 339-346.
- [81] Bergeret L, Black D, Theunis J, Misery L, Chauveau N, Aubry F, Gros H, Viallard G and Celsis P. Validation of a model of itch induction for brain positron emission tomography studies using histamine iontophoresis. *Acta Derm Venereol* 2011; 91: 504-510.
- [82] Valet M, Pfab F, Sprenger T, Woller A, Zimmer C, Behrendt H, Ring J, Darsow U and Tolle TR. Cerebral processing of histamine-induced itch using short-term alternating temperature modulation—an FMRI study. *J Invest Dermatol* 2008; 128: 426-433.
- [83] Herde L, Forster C, Strupf M and Handwerker HO. Itch induced by a novel method leads to limbic deactivations a functional MRI study. *J Neurophysiol* 2007; 98: 2347-2356.

Neural processing of pain and itch

- [84] Ishiiji Y, Coghill RC, Patel TS, Oshiro Y, Kraft RA and Yosipovitch G. Distinct patterns of brain activity evoked by histamine-induced itch reveal an association with itch intensity and disease severity in atopic dermatitis. *Br J Dermatol* 2009; 161: 1072-1080.
- [85] Mochizuki H, Inui K, Tanabe HC, Akiyama LF, Otsuru N, Yamashiro K, Sasaki A, Nakata H, Sadato N and Kakigi R. Time course of activity in itch-related brain regions: a combined MEG-fMRI study. *J Neurophysiol* 2009; 102: 2657-2666.
- [86] May AC, Stewart JL, Tapert SF and Paulus MP. The effect of age on neural processing of pleasant soft touch stimuli. *Front Behav Neurosci* 2014; 8: 52.
- [87] Papoiu AD, Emerson NM, Patel TS, Kraft RA, Valdes-Rodriguez R, Nattkemper LA, Coghill RC and Yosipovitch G. Voxel-based morphometry and arterial spin labeling fMRI reveal neuropathic and neuroplastic features of brain processing of itch in end-stage renal disease. *J Neurophysiol* 2014; 112: 1729-1738.
- [88] Leknes SG, Bantick S, Willis CM, Wilkinson JD, Wise RG and Tracey I. Itch and motivation to scratch: an investigation of the central and peripheral correlates of allergen- and histamine-induced itch in humans. *J Neurophysiol* 2007; 97: 415-422.
- [89] Papoiu AD, Coghill RC, Kraft RA, Wang H and Yosipovitch G. A tale of two itches. Common features and notable differences in brain activation evoked by cowhage and histamine induced itch. *Neuroimage* 2012; 59: 3611-3623.
- [90] Papoiu AD, Nattkemper LA, Sanders KM, Kraft RA, Chan YH, Coghill RC and Yosipovitch G. Brain's reward circuits mediate itch relief. a functional MRI study of active scratching. *PLoS One* 2013; 8: e82389.
- [91] Descalzi G, Chen T, Koga K, Li XY, Yamada K and Zhuo M. Cortical GluK1 kainate receptors modulate scratching in adult mice. *J Neurochem* 2013; 126: 636-650.